

The Effectiveness of Organotin (IV) Benzylisopropylthiocarbamate Compounds as Insecticide Against *Aedes Aegypti* Linn (Diptera: Culicidae) in Laboratory

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Abstract: Problem statement: The widespread use of insecticides had resulted in insecticide resistance of vectors of dengue as well as polluting the environment. Organotin (IV) compounds had the potential to be developed as the insecticides to overcome the existing problem. **Approach:** The aim of this study was to examine the insecticidal effects which were larvicidal and adulticidal effects of organotin (IV) benzylisopropylthiocarbamate compounds against *Aedes Aegypti* Linn. In laboratory larvicidal bioassay test of a series of three organotin (IV) benzylisopropylthiocarbamate compounds on third instar larvae of *Aedes Aegypti* had been carried out. **Results:** The study was found that compound B showed the best larvicidal effect with the LC₅₀ and LC₉₀ values of 0.004 ppm and 0.007 ppm, respectively. Compound C was also displayed good larvicidal effect with the LC₅₀ and LC₉₀ values of 0.029 ppm and 0.108 ppm, respectively. While, compound A was shown the least larvicidal effect with the LC₅₀ and LC₉₀ values of 0.404 ppm and 0.749 ppm, respectively. Further testing was conducted on compound B on adults of *Aedes aegypti* female to investigate its adulticidal property. The result showed that compound B displayed good adulticidal activity with LC₅₀ dan LC₉₀ of 4.277 ppm and 27.653 ppm, respectively. **Conclusion:** Compound B is the most effective compound among three organotin (IV) benzylisopropylthiocarbamate compounds tested against the dengue vector *Aedes Aegypti* and has potential to be explored as an insecticide to control the spread of dengue fever.

Key words: Dithiocarbamates, benzylisopropylthiocarbamate, adulticidal activity, *aedes aegypti*, organotin (IV), mosquito, larvicidal activity, larvae,

INTRODUCTION

Dengue fever is a mosquito-borne disease that threatens international public health. The disease is found in many tropical and subtropical countries throughout the world, especially in urban areas and semi urban (WHO, 2007). Dengue fever is caused by dengue virus. There are four dengue viruses stereotype known as DEN-1, DEN-2, DEN-3 and DEN-4 (WHO, 1999). *Aedes Aegypti* and *Aedes albopictus* is the main vector of this disease, but the *Aedes Aegypti* is the most efficient vector of dengue fever since it live in the house and surrounding houses (Rohani *et al.*, 1997).

Until now no vaccine is completely appropriate to be given to society for the prevention of further spread

of dengue fever (Chapagain *et al.*, 2008). Thus, vector control is the most effective method of controlling the spread of the disease (Zaim and Guillet, 2002) and chemical insecticide is a method that is believed to reduce the *Aedes* population and as the result incidence of dengue fever can be reduced (Choocote *et al.*, 2006).

Unfortunately, the uncontrolled use of insecticide has resulted in problems of insecticide resistance (Somboon *et al.*, 2003). In addition, the effect of degradation of a toxic insecticide metabolites in the environment leading to environmental pollution such as pollution of surface water and ground water (Dua *et al.*, 2010) as well as negative effects on non-target organisms, including

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human (Yang *et al.*, 2002). Recognizing this problem, scientists seek new alternatives in the production of environmentally friendly insecticide and at the same time preventing resistance in the vector.

Wide application of organotin (IV) compounds in the industry (Boraiko and Batt, 2005) and its potential in biological activity has attracted researchers to conduct studies on this compound (Wiecek *et al.*, 2010). Among the biological activities of organotin (IV) compound have these compounds identified as a potential antiviral agent (Sreeja and Kurup, 2005), antineoplastic agents, antituberculosis (Gielen, 2002; Dokorou *et al.*, 2004; Kovala-Demertzi *et al.*, 2005; Kovala-Demertzi, 2006), antibacterial agent (Jamil *et al.*, 2009), anticancer agent (Awang *et al.*, 2010) and so on.

Use of organotin (IV) compounds in the field of entomology is also now getting attention (Shukla *et al.*, 2010). Several studies showed organotin (IV) compounds were effective against some pests like periplanata Americana, musca domestica, Spodoptera litura and Tetranychus urticae (Shukla *et al.*, 2010) and a few species of larvae and adult mosquitoes like *Aedes Aegypti* and *Aenopheles stephensi* (Eng *et al.*, 2003; 2007; Baul *et al.*, 2005; 2010). Group and the nature of organic group that bound to tin atom, such as the monoalkyl, dialkyl and trphenyl are the basic factors determining the biological activity (Boraiko and Batt, 2005; Awang *et al.*, 2010; Pellerito *et al.*, 2006).

Until now there has been reports of *Ae. aegypti* resistance to organotin (IV) compound and reported that this compound will be degraded to non-toxic inorganic compound in the environment (Blunden and Chapman, 1986; Buck-Koehntop *et al.*, 2006).

In this study, we report the insecticidal activity of three compounds of organotin (IV) bezylisopropylidithiocarbamate that were dimethyltin (IV) (compound A), dibutyltin (IV) (compound B) and triphenyltin (IV) bezylisopropylidithiocarbamate (compound C) against *Ae. aegypti* mosquito in laboratory.

MATERIALS AND METHODS

Materials: Organotin (IV) bezylisopropylidithiocarbamate compounds (A-C).

Larvae and adult mosquito: The *Aedes Aegypti* mosquito larvae and adult mosquitoes were obtained from the colonies that had been reared continuously for generations in a laboratory free of exposure to pathogens and insecticides. They were maintained at 25-30°C and 80-90% relative humidity under a photoperiod of 12:12 h (light/dark) in the Insectarium of the Program in Biomedical Science, Faculty of

Health Sciences, Universiti Kebangsaan Malaysia. The larvae were fed with a ground beef liver that have been dried and grind after reaching the 1st instar. The dechlorinated water that contains the larvae and the beef liver must be changed regularly to ensure the water condition is always clean and clear. The adults were reared in cages and were provided with 10% sucrose added with multivitamins. Female mosquitoes were periodically blood-fed to restrained guinea pigs to obtain protein used principally for egg production. Under these conditions, the full development from egg to adult lasted about 3-4 weeks. Batches of 2-5 day-old healthy female mosquitoes were used in the adulticidal bioassay.

Preparation of the organotin (IV) bezylisopropylidithiocarbamate stock solutions: Stock solutions of the organotin(IV) bezylisopropylidithiocarbamate were prepared in either 95% ethanol, Dimethyl Sulfoxide (DMSO) or acetone depending on the solubility of the compound at concentration 100 parts per million (ppm). The dissolution of the organotin (IV) bezylisopropylidithiocarbamate in the organic media was to facilitate the dispersion of the compounds in water. The acetone and DMSO were spectrograde quality while the 95% ethanol was reagent grade.

Larvicidal bioassay testing: The larvicidal bioassay testing was followed the method from Baul *et al.* (2010) with slight modification. This testing was performed in 30 ml disposable cups using ten larvae of *Ae. aegypti* in the 3rd instar stage. Solution of compounds 1-3 were added to 15 ml of distilled water that had been prepared in a disposable cup. The *Ae. aegypti* larvae were then transferred into the solution and distilled water to give the desired concentration of solution. The total assay volume in each case was 20 mL. A solution containing distilled water and DMSO, but without the organotin (IV) solution, served as a negative control and temephos was used as a positive control. Mortalities were recorded at 24 h of exposure. The moribund and dead larvae in three replicates were combined and expresses as a percentage mortality of each concentration. The larvae were considered dead if they showed no sign of swimming movements even after probing with a needle.

Adulticidal bioassay testing: Further testing on the adulticidal activity of the most effective compound in the larvicidal bioassay testing was conducted on adults of *Ae. aegypti* female. The adulticidal bioassay testing was performed by topical application of the compound, following slightly modified versions of the WHO standard protocols (WHO, 2006). The Adulticidal activity of the compound was evaluated at

four concentrations and 25 females were used for each concentration of the most effective compound. Non-blood-fed female mosquitoes (2-5 days old) were briefly anesthetized with extreme temperature for 30 seconds and placed on a cold plate. The selected compound solution (1 µL) dissolved in DMSO was applied onto the upper part of the immobilized mosquito's pronotum using multipette plus (Eppendorf Research, Jerman model no. 4981950). A solution containing distilled water and DMSO, but without the compound solution, served as a negative control and malathion was used as a positive control for comparison. Both groups were treated in a similar manner to that treated group. After each test, females were transferred into disposable cups and 10% sucrose solutions with added multivitamin on cotton wools were provided. After application, the females in all groups were maintained at 27±3°C and 80±10% RH in plastic cups. At the end of a 24-h recovery period, the mosquitoes were considered dead if they showed no sign of movement such as lying on the bottom of the plastic cup and not responding to mechanical stimulation. Three replicates were carried out with mosquitoes from different rearing batches and the results were pooled.

Statistical analysis of data: Tests with more than 20% control mortality were discarded and the repeated. However, if the mortality of control was between 5-20%, the observed percentage mortality was corrected by Abbot's formula (Abbott, 1925).

$$\% \text{ mortality} = \frac{\% \text{ test mortality} - \% \text{ control mortality}}{100 - \% \text{ control mortality}} \times 100$$

LC₅₀ and LC₉₀ with their 95% confidence limits of the compound was determined using computerized Log probit analysis test (Raymond, 1985).

RESULTS

Larvicidal bioassay testing: The LC₅₀ and LC₉₀ values expressed in parts per million and their 95% confidence limit for compounds A-C screened against the third instar larvae of *Ae. aegypti* mosquitoes are given in Table 1. As can be seen from Table 1, compounds A, B and C had the good larvicidal activity on third instar larvae of *Ae. aegypti* mosquitoes because all LC₅₀ and LC₉₀ values for these three compounds were less than 1 ppm. The data showed that there were differences in larvicidal effect of compounds which compound B displayed the best larvicidal effect with the LC₅₀ and LC₉₀ values of 0.004 and 0.007 ppm, respectively.

Table 1: Lethal concentration of compound A-C and temephos against third instar *Ae. aegypti* mosquito larvae after 24 h exposure

Compound	LC ₅₀ (ppm) (Confidence interval 95%)	LC ₉₀ (ppm) (Confidence interval 95%)	Gradient
A	0.404 (0.377-0.438)	0.749 (0.671-0.867)	4.779±8.473
B	0.004 (0.004-0.005)	0.007 (0.007-0.009)	5.822±0.580
C	0.029 (0.024-0.034)	0.108 (0.086-0.147)	2.240±0.207
Temephos	0.011 (0.010-0.013)	0.026 (0.021-0.036)	3.483±1.180

Table 2: Lethal concentration of compound B and malathion against adults of *Ae. aegypti* mosquito after 24 h exposure

Compound	LC ₅₀ (ppm) (Confidence interval 95%)	LC ₉₀ (ppm) (Confidence interval 95%)	Gradient
B	4.277 (3.221-6.377)	27.653 (15.318-72.478)	1.581±0.204
Malathion	4.368 (3.313-6.597)	38.310 (19.408-124.021)	1.359±0.191

Compound C showed a good larvicidal effect with the LC₅₀ and LC₉₀ values of 0.029 ppm and 0.108 ppm, respectively while the least larvicidal effect showed by compound A with the LC₅₀ and LC₉₀ values of 0.404 and 0.749 ppm, respectively.

Adulticidal bioassay testing: The result of adulticidal bioassay testing was summarized in Table 2. The result showed compound B also had good adulticidal effective against adults of *Ae. aegypti* mosquito with LC₅₀ and LC₉₀ values of 4.277 and 27.653 ppm, respectively.

DISCUSSION

From the results, the order of larvicidal activity based on the organic group attached to the tin atom can be observed which compound that attached to a dibutyl group (compound B) was the most effective larvicide, followed by a compound that attached with triphenyl group (compound C) and compound that attached to a dimethyl group (compound A) to the tin atom. However, when compared to the LC₅₀ and LC₉₀ values of temephos which was the gold standard larvicidal testing, only compound B could equal the effectiveness of temephos as a larvicide against *Ae. aegypti* mosquito larvae while the effectiveness of compound A and C as larvicide against *Ae. aegypti* mosquito larvae is less than temephos.

When compared with other larvicidal activity studies of organotin (IV) compounds, these compounds which used in this study were among organotin (IV) compounds that had a good larvicidal effect. The range

of LC₅₀ values of triorganotin 2-(*p*-chlorophenyl)-3-methylbutyrate was 0.32-3.13 ppm (Eng *et al.*, 2007). This value indicated that compound B and C were more effective as larvicide against *Ae. aegypti* mosquito larvae than these compounds. For triphenyltin *para*-substituted benzoate compound and tricyclohexyltin *para*-substituted benzoate compound, the average of the LC₅₀ values were 0.62 and 1.16 ppm, respectively (Duong *et al.*, 2006). Based on the LC₅₀ values, compound A-C had more potential to be explored as larvicide on *Ae. aegypti* mosquito larvae compared to triphenyltin and tricyclohexyltin *para*-substituted benzoate compounds. Larvicidal effects of compounds A-C on *Ae. aegypti* mosquito larvae had also been better than triorganotin (IV) complex compound because compound A-C have higher LC₅₀ values (Baul *et al.*, 2005). Organic group and ligand that attached to the tin atom were the main factor that influence the biological activity of organotin (IV) (Boraiko and Batt, 2005; Awang *et al.*, 2010; Pellerito *et al.*, 2006). However, further study should be conducted to clarify the mechanism of toxicity of compound A-C to the *Ae. aegypti* mosquito larvae.

From the larvicidal bioassay testing, it was shown that compound B performed the best larvicidal activity against third instar *Ae. aegypti* mosquito larvae compared to compound A and C. Therefore, further study on compound B on adults of *Ae. aegypti* mosquito was conducted to investigate its adulticidal property by a WHO topical application method. The purpose of this method was to determine the intrinsic activity of this compound of a target species to isolate toxicity of from the confounding effects resulting from insect behavior (WHO, 2006). Remarkably, the effectiveness of the compound B was comparable to the effectiveness of malathion which was the gold standard for mosquito adulticide.

The advantages of organotin (IV) compound are it will biodegrade into a non-toxic inorganic compound in the environment and there is no reported resistance of this compound towards *Ae. aegypti*. With these advantages, these organotin (IV) compounds have potential to be explored as an insecticide in the effort to control the widespread of dengue.

CONCLUSION

In conclusion, compound B is the most effective compound among three organotin (IV) compound tested against *Ae. aegypti*. The larvicidal effect of the compound towards *Ae. aegypti* mosquito larvae is superior to temephos while its adulticidal effect towards adults of *Ae. aegypti* mosquito was comparable with malathion. Further study of this

compound should be conducted in order to explore its potential as an insecticide to control the dengue vectors in addition to control the spread of dengue.

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REFERENCES

- Abbott, W.S., 1925. A method of computing the effectiveness of an insecticide. *J. Econ. Entomol.*, 18: 265-266.
- Awang, N., I. Baba, N.S.A. Mohd Yusof and N.F. Kamaludin. 2010. Synthesis and characterization of organotin (IV) N-Benzyl-N-Isopropylthiocarbamate compounds: Cytotoxic assay on human hepatocarcinoma cells (HepG2). *Am. J. Applied Sci.* 7: 1047-1052.
- Baul, T.S.B., K.S. Singh, M. Holcapek, R. Jirasko and A. Linden *et al.*, 2005. Electrospray ionization mass spectrometry of tributyltin(IV) complexes and their larvicidal activity of mosquito larvae: crystal and molecular structure of polymeric (Bu₃Sn[O₂CC₆H₄{N[DOUBLE BOND]N(C₆H₃-4-OH(C(H)[DOUBLE BOND]NC₆H₄OCH₃-4))}-o])_n. *Applied Organomet. Chem.*, 19: 935-944. DOI: 10.1002/aoc.937
- Baul, T.S.B., P. Das, E. Rivalora, X. Song and G. Eng, 2010. Synthesis, Spectroscopic Characterization and Structures of Tributyltin(IV) 4-(((E)-1-(2-hydroxy-5-[(E)-2-(aryl)-1-diazenyl]phenyl)methylidene)amino]benzoates. Toxicity Studies on the Second Larval Instar of the Anopheles stephensi Mosquito Larvae. *J. Inorg. Organomet. Polym.*, 20: 61-68. DOI: 10.1007/s10904-009-9308-2
- Blunden, S.J. and A. Chapman, 1986. In *Organometallic Compounds in the Environment*. 1st Edn., Wiley and Sons, New York.
- Boraiko, C. and J. Batt, 2005. Evaluation of employee exposure to organic tin compounds used as stabilizers at PVC processing facilities. *J. occup. environ. Hyg.* 2: 73-76. PMID: 15764527
- Buck-Koehntop, B.A., F. Porcelli, J.L. Lewin, C.J. Cramer and G. Veglia, 2006. Biological chemistry of organotin compounds: Interactions and dealkylation by dithiols. *J. Organic. Chem.*, 69: 1748-1755. DOI: 10.1002/chin.200628275

- Chapagain, B.P., V. Saharan and Z. Wiesman, 2008. The Larvicidal activity of saponins from *Balanites aegyptiaca* callus against *Aedes Aegypti* mosquito. *Bioresou. Technol.*, 99: 1165-1168. DOI: 10.1016/j.biortech.2007.02.023
- Choocote, W., U. Chaitong, K. Kamsuk, E. Rayyanachanpichal and A. Jitpakdi *et al.*, 2006. Adulticidal activity against *Stegomyia aegypti* (Diptera: Culicidae) of three *Piper spp.* *Rev. Inst. Med. Trop Sao Paulo.*, 48: 33-37. PMID: 16547577
- Dokorou, V., D. Koyala-Demertzi, J.P. Jasinski and A. Galani *et al.*, 2004. Synthesis, Spectroscopic Studies and Crystal Structures of Phenylorganotin Derivatives with [Bis(2,6-dimethylphenyl)amino]benzoic Acid: Novel Antituberculosis Agents. *Helvetica Chimica. Acta.* 87: 1940-1950. DOI: 10.1002/hlca.200490175
- Dua, V.K., A.C. Pandey and A.P. Dash, 2010. The Adulticidal activity of essential oil of *Lantana camara* leaves against mosquitoes. *Indian J. Med. Res.*, 131: 434-439. PMID: 20418559
- Duong, Q., X. Song, E. Mitrojjorgji, S. Gordon and G. Eng, 2006. Larvicidal and structural studies of some triphenyl- and tricyclohexyltin para-substituted benzoates. *J. Organometallic Chem.*, 691: 1775-1779. DOI: 10.1016/j.jorganchem.2005.12.005
- Eng, G., X. Song, A. Zapata, A.C.D. Dios and L. Casabianca *et al.*, 2007. Synthesis, structural and larvicidal studies of some triorganotin 2-(p-chlorophenyl)-3-methylbutyrates. *J. Organometallic Chem.*, 692: 1398-1404. DOI: 10.1016/j.jorganchem.2006.11.030
- Eng, G., X. Song, Q. Duong, D. Strickman and J. Glass *et al.*, 2003. Synthesis, structure characterization and insecticidal activity of some triorganotin dithiocarbamates. *Applied Organometallic Chem.*, 17: 218-225. DOI: 10.1002/aoc.423
- Gielen, M., 2002. Review: Organotin compounds and their therapeutic potential: A report from the organometallic chemistry department of the free university of Brussels. *Applied Organometallic Chem.*, 16: 481-494. DOI: 10.1002/aoc.331
- Jamil, K., M. Bakthiar, A.R. Khan, F. Rubina and R. Rehana *et al.*, 2009. Synthesis characterization and antimicrobial activities of novel organotin compounds. *Afr. J. Pure Applied Chem.*, 3: 60-65.
- Kovala-Demertzi, D., 2006. Recent advances in non-steroidal anti-inflammatory drugs, NSAIDs: Organotin complexes of NSAIDs. *J. Organometallic Chem.*, 691: 1767-1774. DOI: 10.1016/j.jorganchem.2005.11.058
- Kovala-Demertzi, D., V.N. Dokorou, J.P. Jasinski, A. Opolski and J. Wiecek *et al.*, 2005. Organotin flufenamates: Synthesis, characterization and antiproliferative activity of organotin flufenamates. *J. Organometallic Chem.*, 690: 1800-1806. DOI: 10.1016/j.jorganchem.2005.02.004
- Pellerito, C., L. Nagy, L. Pellerito and A. Szorcisk, 2006. Biological activity studies on organotin (IV) n+ complexes and parent compounds. *J. Organometallic Chem.*, 691: 1733-1747. DOI: 10.1016/j.jorganchem.2005.12.025
- Raymond, M., 1985. Log-probit analysis basic program of microcomputer. *Cashiers ORSTOM series Entomologie MED. Parasitologie*, 22: 117-121.
- Rohani, A., W.A. Nazni, L.V. Ngo, I. Ibrahim and H.L. Lee, 1997. Adulticidal properties of the essential extracts of some Malaysian plants on vector mosquitoes. *Tropical Biomed.* 14: 5-9.
- Shukla, S.K., V.K. Tiwari, S. Rani, K. Ravi and I.C. Tewari, 2010. Studies on insecticidal and pesticidal activity of some organotin compounds. *Inter. J. Agricul. Sci.*, 2: 5-10.
- Somboon, P., L.A. Prapanthadara and W. Suwonkerd, 2003. Insecticide susceptibility tests of *Anopheles minimus s.l.*, *Aedes aegypti*, *Aedes albopictus* and *Culex quinquefasciatus* in northern Thailand. *Southeast Asian J. Trop. Med. Public Health*, 34: 87-93. PMID: 12971519
- Sreeja, B.P. and M.R. Kurup, 2005. Synthesis and spectral characterization of ternary complexes of oxovanadium (IV) containing some acid hydrazones and 2,2'-bipyridine. *Spectrochim Acta* 61: 331-336. PMID: 15556457
- WHO, 1999. Prevention and control dengue and dengue haemorrhagic fever: Comprehensive guidelines. WHO Regional Publication.
- WHO, 2006. Guidelines for testing mosquito adulticides for indoor residual spraying and treatment of mosquito nets. World Health Organization.
- WHO, 2007. Global Insecticide Use for Vector-borne Disease Control. 3rd Edn., World Health Organization, Geneva, pp: 81.
- Wiecek, J., D. Kovala-Demertzi, Z. Ciunik, J. Wietrzyk and M. Zervou *et al.*, 2010. Organotin compound derived from 3-hydroxy-2-formylpyridine semicarbazone: synthesis, crystal structure and antiproliferative activity. *Bioinor. Chem. Appli.* PMID: 20490260
- Yang, Y.C., S.G. Lee, H.K. Lee, M.K. Kim and H.S. Lee, 2002. A piperidine amide extracted from *Piper longum* L. Fruit shows activity against *Aedes Aegypti* mosquito larvae. *J. Agric. Food Chem.*, 50: 3765-3767. DOI: 10.1021/jf011708f
- Zaim, M. and P. Guillet, 2002. Alternative insecticides: An urgent need. *Trends Parasitol.*, 18: 161-163. DOI: 10.1016/S1471-4922(01)02220-6