

Development of Three Bacteria Consortium for the Bioremediation of Crude Petroleum-oil in Contaminated Water

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Abstract: We have developed active microbial consortium that could be of higher degradation of crude oil contaminated groundwater, wastewater aeration pond and biopond at the oil refinery Terengganu Malaysia. Among four isolates that showed good growth only three different isolates (*Acinetobacter faecalis* WD2, *Staphylococcus*. sp DD3 and *Neisseria elongate* TDA4.) were selected based on the growth ability and degradation. Significant growth and effectiveness of hydrocarbon biodegradation of the bacterial consortium examined bacterial strains and their mixtures in both were observed after 5, 10 and 15 days of degradation. Gas chromatography showed that more than 96 and 98% degradation of total hydrocarbon by consortia sp respectively.

Key words: Biodegradation, crude oil contaminated water

INTRODUCTION

Chemical treatment includes direct injection of chemical oxidants into contaminated soil and groundwater thereby altering native aquatic chemistry and biology. Biological treatment most commonly involves the breakdown of contamination into nontoxic forms using microbiological processes^[1]. Thus, bioremediation may be defined as the use of living organisms to remove environmental pollutants from soil, water and gases^[2]. The advantages of employing mixed cultures as opposed to pure cultures in bioremediation have been widely demonstrated. It could be attributed to the effects of synergistic interactions among members of the association. The mechanisms in which petroleum degraders' benefit from synergistic interactions may be complex. It is possible that one species removes the toxic metabolites (that otherwise may hinder microbial activities) of the species preceding it. It is also possible that the second species are able to degrade compounds that the first are able to only partially^[3]. Further research should be directed towards understanding the roles of individual members in influencing the effectiveness of a microbial consortium. Demonstrated a consortium of 8 strains made up of members of 6 genera to be able to effectively degrading crude oil^[4]. Interestingly, only 5 of these strains were able to grow in pure cultures using

a variety of hydrocarbons. However, when the other 3 strains were removed from the consortium, the effectiveness of the mixed culture was remarkably reduced. These further support the theory that each member in a microbial community has a significant role and may need to depend on the presence of other species or strains to be able to survive. They are produced by many bacterial strains that can degrade or transform the components of petroleum products. They are non-toxic, non-hazardous, biodegradable and environmentally friendly compounds^[5].

MATERIALS AND METHODS

Source of microorganisms, media and culture condition: Bacterial strains were isolated from samples collected from groundwater and wastewater aeration pond and biopond located at the Terengganu oil refinery, Malaysia. 10ml of each sample were washed with 90mL saline and filtered with membrane. Incubation was carried out for 24h at 37°C on nutrient agar plate. Mineral Salts Medium (MSM)^[6] was prepared by dissolving 1.8g K₂HPO₄, 4.0g NH₄Cl, 0.2g MgSO₄·7H₂O, 0.1g NaCl, 0.01g FeSO₄·7H₂O in 1L of distilled water. Bacteriological agar was added (15 g/l) to the solution where solid basal medium was required. The pH was adjusted to 6.90 and the medium was autoclaved at 121°C for 15 min. 1.0%(v/v) tapis crude

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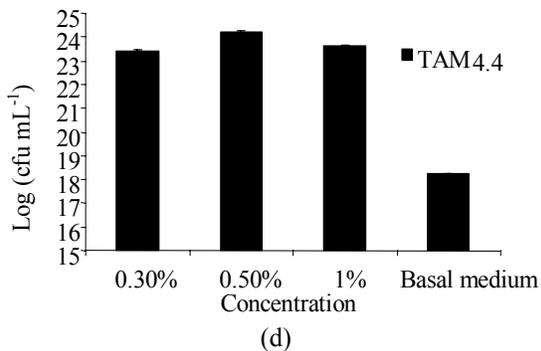
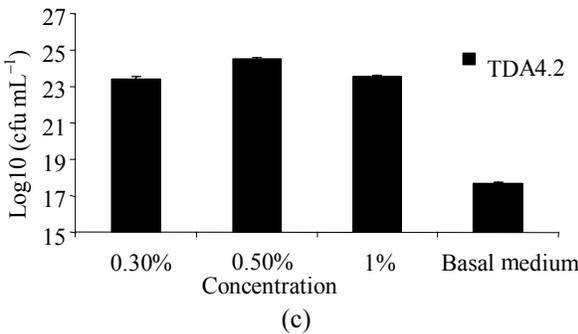
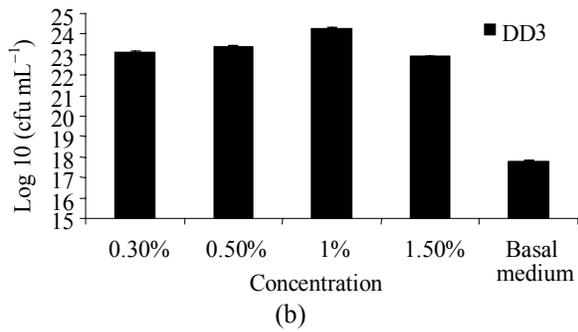
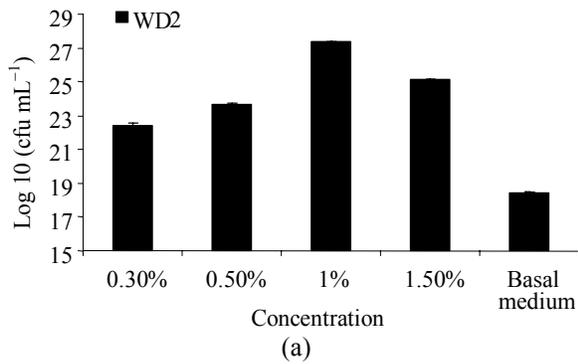


Fig. 1: Effect of different concentration of peptone on growth of isolates (a) *Acinetobacter faecalis* (b) *Staphylococcus. sp* (c) *Neisseria elongate* and (d) , *Pseudomonas putida*

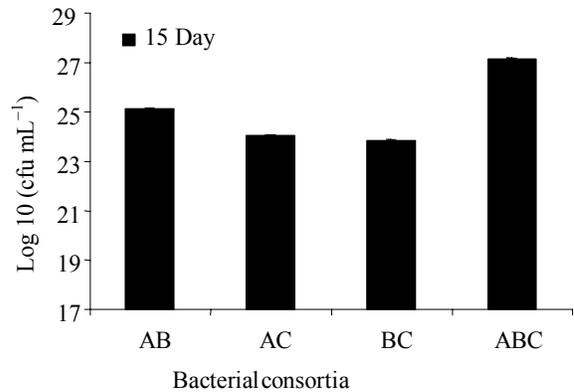
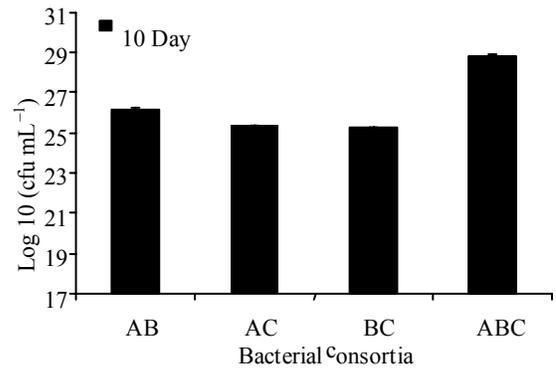
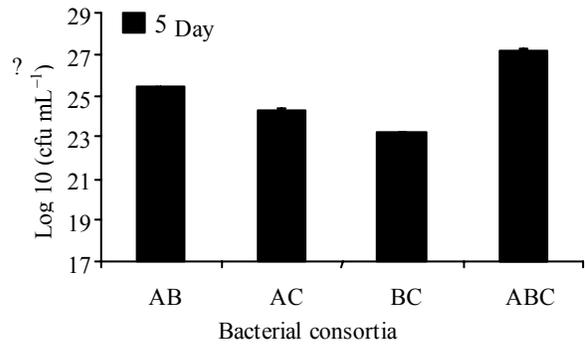


Fig.2: Growth of isolates (WD2, DD3, and TDA4.2) as consortium throughout 5, 10 and 15 days in the MSM medium with 1% Tapis crude oil

oil was added as the sole carbon source and complex vitamin solution (1.0 mL⁻¹) according to manufacturer's suggestion.

Effect of nitrogen source on growth and biodegradation: To determine the effect of the organic nitrogen source on biodegradation, the inorganic nitrogen sources of the basal medium were substituted by the addition of the following; peptone at

concentrations of 0.30, 0.50, 1 and 1.50% (v/v). Cultivations were carried out after 5 days and the growth was measured as CFU count

Construction of bacterial consortia: The three different isolates that demonstrated good growth were chosen to construct consortia of hydrocarbon degraders. In total, three different bacteria were constructed consortia and tested for in this study to confirm their biodegradation capabilities.

Preparation of consortia inoculum: The isolates were grown separately in NB and processed to yield separate suspensions with an absorbance reading of 0.5 at 550 nM. Specific aliquots of the bacterial inoculum were then separately added into normal saline solution to give a final combined inoculum concentration of 10% (v/w).

Extraction of residual crude oil for Gas Chromatography (GC) analysis: Crude oil was extracted with a pre-cleaned separating funnel, following the modified methods of^[7]. 50 mL of sample (BM + crude oil + bacteria), were then centrifuged at 4000 rpm for 15 min. The separating funnel was stoppered with a glass stopper and it was shaken vigorously. Vapour was carefully vented out through the stopcock. The process of venting and vigorous shaking was repeated for several minutes and the mixture was allowed to separate into two phases. After phases have been separated in the funnel, chloroform was collected in a 250 mL pre-cleaned bottle. The extraction was repeated with 25 mL chloroform twice. Then, it was filtered and dried by passing it through of 3g pre-combusted Na₂SO₄ (150°C for 3h in incubation) and collected in a round bottom flask. The extract was then evaporated by using a rotary evaporator. The crude oil extracted was washed with 2 mL chloroform. The solvents were removed under a gentle stream (dried in fume chamber) for 7 days. The residue was reconstituted with 1mL of chloroform and analyzed by GC.

Analyses of extract: A detailed analysis of the hydrocarbon extract was performed by Gas Chromatography GC. The clarus GC 500 was equipped with a split injector (split ratio 50/1) and a Flame Ionization Detector FID both set at 300°C; gas carrier was nitrogen 1.50 mL min⁻¹; the column was fused silica capillary column (30.0 mx0.32 mm, film thickness 0.25µm); temperature programming was 60-320°C, 5°C min⁻¹, injection volume 1µL.

Table1: Three bacteria consortium selected for biodegradation

Bacteria	Consortium
<i>Acinetobacter faecalis</i> and <i>Staphylococcus</i> . sp	A+B
<i>Acinetobacter faecalis</i> and <i>Neisseria elongate</i>	A+C
<i>Staphylococcus</i> . sp and <i>Neisseria elongate</i>	B+C
<i>Acinetobacter faecalis</i> , <i>Staphylococcus</i> . sp and <i>Neisseria elongate</i>	A+B+C

RESULTS AND DISCUSSIONS

Effect of peptone on growth and degradation: The bacterial growths in different percentages of peptone as nitrogen source were studied (Fig. 1). All four isolates showed good growth at different peptone concentrations 0.3, 0.5, 1.0 and 1.50%. Optimum growths were observed at percentages 1% Tapis crude oil on the 5 day of incubation. Among the four isolates, isolate WD2 and DD3 showed a maximum growth with 1% peptone while, isolates TDA4.2 and TAM 4.4 showed a maximum growth with 0.5% peptone. From this study, 1% peptone was the best nitrogen source tested for growth compared to the basal medium. From four selected isolates, only three isolate showed good selected for consortium (Table 1). However, increasing the concentration of peptone from 1-1.50% significantly decreases growth and degradation. This is in agreement with the finding by^[8] that due to enhanced growth; degradability of oil was better in a medium containing 1% peptone, compared to medium containing either ammonium sulfate or potassium nitrate. Also reported cell growth increase in the medium with 1% peptone for pseudomonas fluorescences FS1^[9].

Evaluation of bacterial consortia: Growth and degradation of 1% tapis crude oil: The growth of three selected isolates as consortia that could effectively remove hydrocarbon pollutants, following 5, 10 and 15 days incubation at 37°C, pH 7 and with agitation at 150 rpm was measured as CFU counts and degradation was measured using GC (Fig. 2). The consortia were analyzed for the extent of biodegradation in medium containing 1% tapis crude oil contaminated water (Fig. 4 and 5).

Consortia A and B that consisted of the WD2 and DD3 showed increased growth in the cell numbers from 243x10²²-178x10²³ CFU mL⁻¹ within 5 and 10 day and the end of the 15 day decreased to 105x10²¹ CFU mL⁻¹ (Fig. 2).

The consortia showed 97.50% degraded found to degraded 100% short-chain and intermediate while was still limited to longer-chain aliphatic compounds

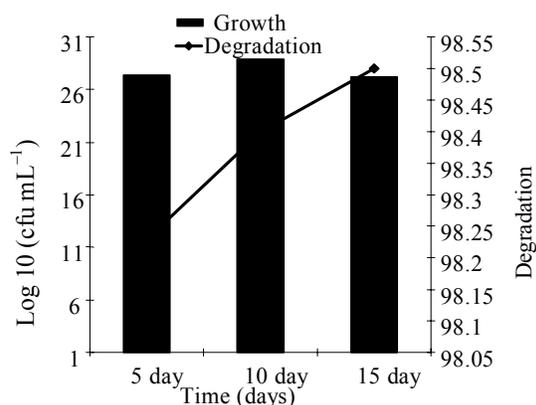


Fig.3: Growth of consortium ABC and degradation at 5, 10, and 15 days in the MSM medium with 1% Tapis crude oil

between 97.56 and 97.52% of C₂₃- C₃₄ remained in the presence of longer-chain between 2.44 and 2.48% of C₂₃ - C₃₄ remained in the present of longer-chain after 5 days (Fig. 4)

The second combination of the two isolates WD2 and TDA2.4 (Mixture A and C) when tested on 1% Tapis crude oil at 5 days of incubation. The cell numbers of strain increased from 148×10^{21} to 254×10^{22} CFU mL⁻¹ within 5 and 10 day at the end of the 15 day study, cell counts decreased to 89×10^{21} CFU mL⁻¹ (Fig. 2). While reduction in amounts of hydrocarbons was detected during gas chromatographic analysis was degraded 96.88% that 100% short-chain and intermediate while was still limited to longer-chain aliphatic compounds between 97.99 and 91.33% of C₂₂ to C₃₅, while between 2.01 and 8.66% remained in the present of longer-chain after 5 days (Fig. 4)

The third consortia of the two isolates DD3 and TDA2.4 (Mixture B and C) when tested on 1% Tapis crude oil at 5 days of incubation. The bacterial cell numbers of strain increased from 156×10^{20} to 188×10^{21} CFU mL⁻¹ within 5 and 10 day at the end of the 15 day study, cell counts decreased to 75×10^{20} CFU mL⁻¹ (Fig.2). The reduction in the amounts of hydrocarbons was detected during gas chromatographic analysis was degraded 96.77% that 100% short-chain and intermediate while was still limited to longer-chain aliphatic compounds between 96.90 and 96.17% of C₂₆ to C₃₇ after 5 days (Fig.4)

The fourfold consortia of three isolates WD2, DD3 and TDA2.4 (Mixture A, B and C), the cell of Consortium A, B and C increased from 135×10^{25} to 175×10^{26} a CFU mL⁻¹ within 5 and 10 day at the end of the 15 day study, cell counts decreased to 134×10^{25} CFU mL⁻¹ (Fig.2).

The higher levels of detected hydrocarbon during gas chromatographic analysis removal were seen with consortium A, B and C 98.23 degraded after 5 days that 100% short-chain, intermediate while was still limited to longer-chain aliphatic compounds between 98.13 and 95.58% of C₂₇ to C₃₆ (Fig.4). further the consortium A, B and C was efficient higher levels of hydrocarbon at degraded 98.25% after 10 days that 100% short-chain, intermediate while was still limited to longer-chain aliphatic compounds between 98.40 and 97.21% of C₂₈ to C₃₄ after 10 days (Fig. 5). While none aliphatic compound degradation was seen higher after 15 day 98.50% degraded between 98.35 and 97.12% of C₂₈ to C₃₃ compared to 10 day (Fig. 3). Consortium mixture A, B and C degraded 100% short-chain and intermediate while was limited to longer-chain aliphatic compounds compared to other consortia.

The bacterial consortium showed that it could degrade up to a maximum of 98% after 15 days incubation. Higher levels (up to 100%) of degraded hydrocarbon were seen in the short-chain and medium chain alkanes compared to the longer chain alkanes. However, after 15 days of incubation post contamination reduction was seen in all the investigated hydrocarbons although the reduction was more significant in the short-chain, medium-chain aliphatic compounds and aromatic compounds compared to longer-chain. This is in agreement since short-chain and medium-chain alkanes are generally easily degraded due to their lower hydrophobicity. Reported that the maximum degradation was achieved of treatment n-alkanes in the range of C₈-C₁₁, which were degraded completely followed by C₁₂-C₂₁, C₂₂-C₃₁ and C₃₂-C₄₀ with degradations percentage of 100, 83-98, 80-85 and 57-73% respectively using bacterial consortium^[10]. In another report by^[11], the biodegradation of n-alkanes C₁₂-C₃₀ and of various aromatic compound in the crude oil was (examined by GC/FID) using *Acinetobacter*. sp T4 culture. The n-alkanes were almost completely degraded, while the aromatic compounds were not. In the culture of *Pseudomonas putida* PB4, neither the n-alkanes nor the aromatic compounds were degraded to any significant degree. In the mixed culture of *Pseudomonas putida* PB4 and *Acinetobacter*. sp. T4, however, both the n-alkanes and aromatic compound were degraded.^[12] Reported that GC profiles after 7 and 18 days showed that biodegradation of hydrocarbons of the crude oil fractions were totally utilized more than that in the control solutions. Reported that the capillary gas chromatographic analysis of the degraded crude oil revealed that crude oil components of chain length C₁₂ to C₃₂ were extensively degraded by *Serratia marcescens* OCS-21 after 16 days of incubation while

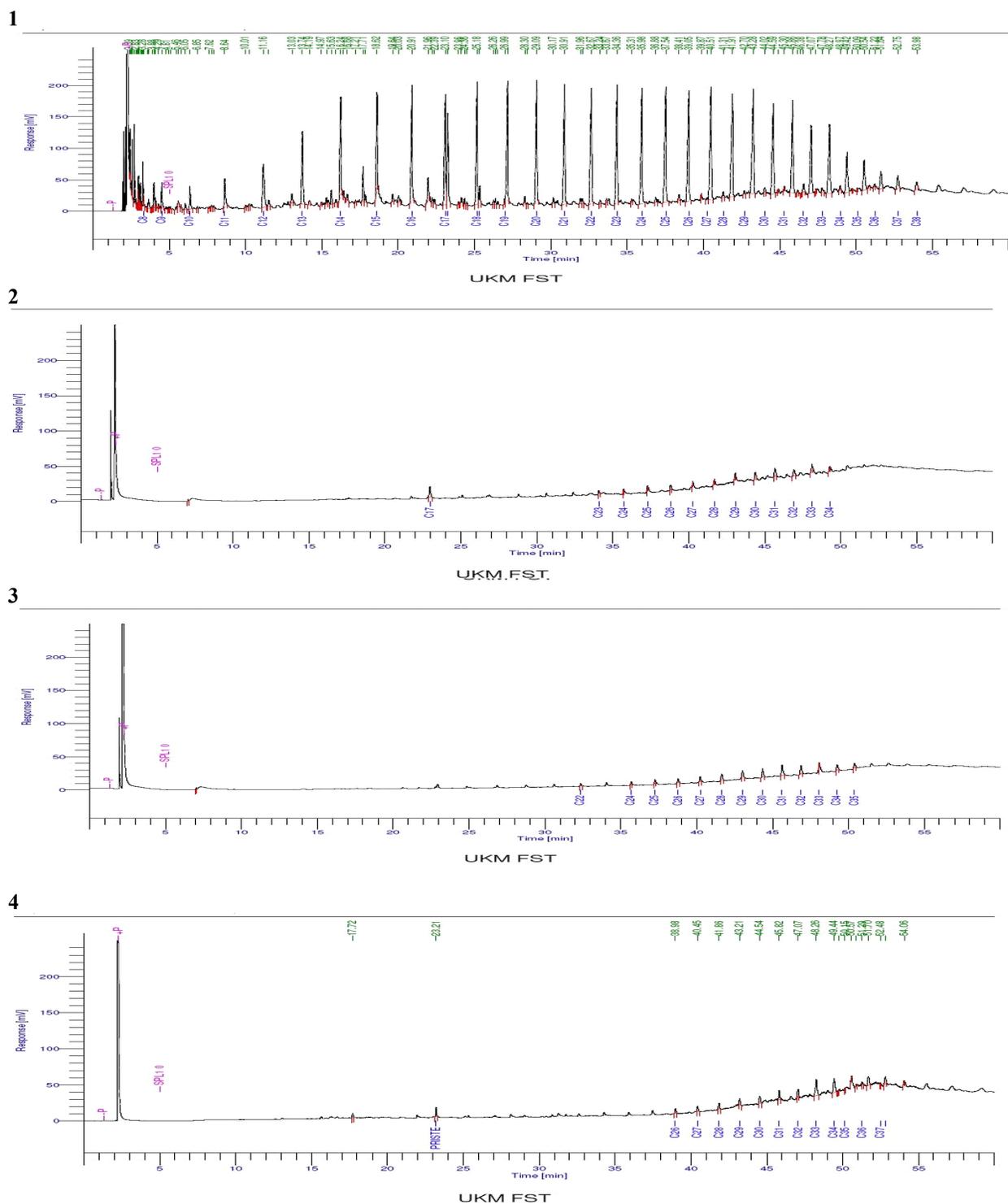
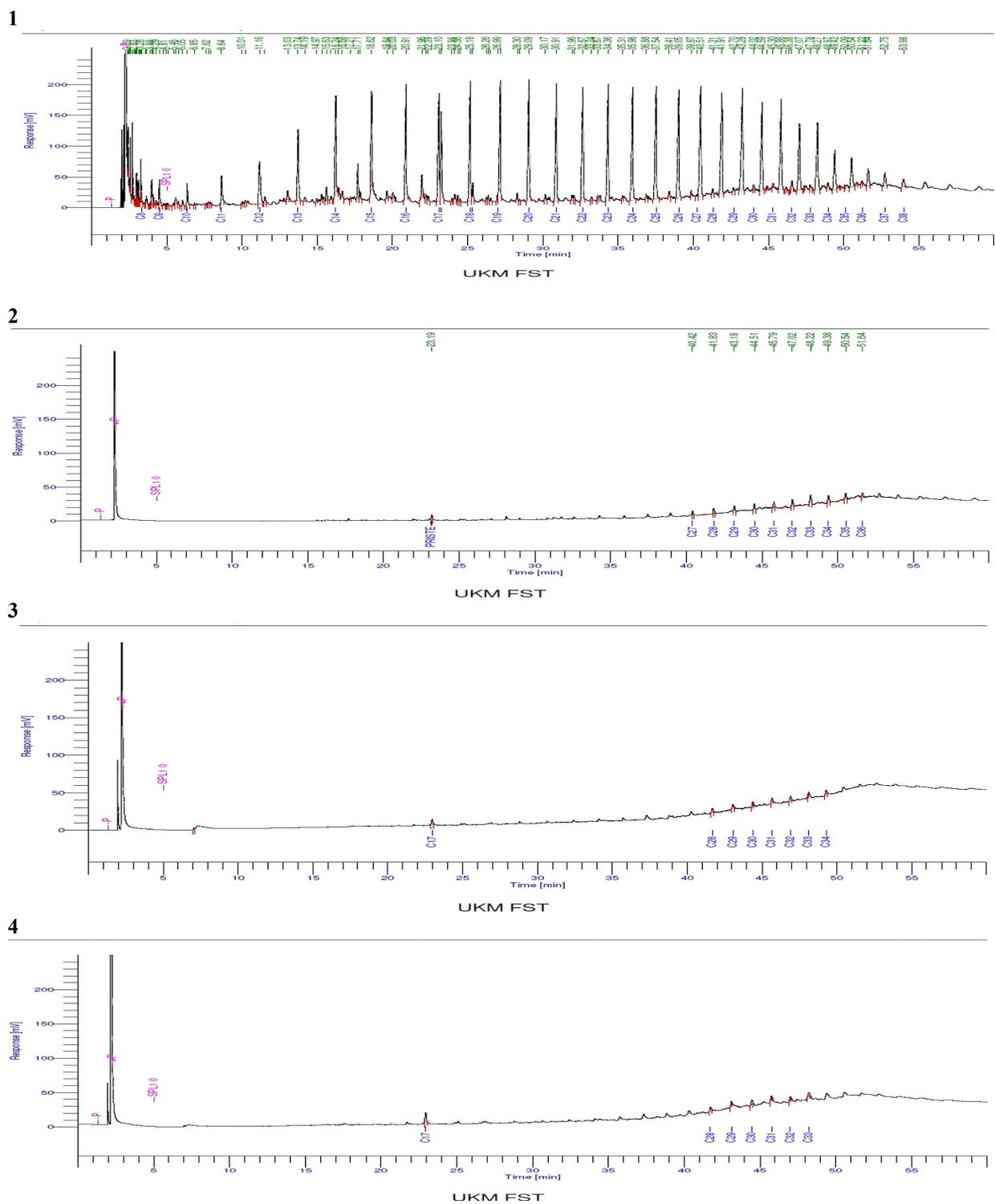


Fig. 4: Chromatogram of 1% Tapis crude oil (1) before microbial degradation (2) degradation by microbial consortium [AB], (3) degradation by microbial consortium [AC], (4) degradation by microbial consortium [BC], after 5 days with MSM medium + peptone



Acinetobacter calcoaceticus COU-27 was able to degrade only C₂₂ to C₃₀ components of the crude oil after 16 days of incubation. All four isolates showed a maximum of hydrocarbon removals which were seen with the short-chain and medium-chain aliphatic compounds compared to longer-chain aliphatic compounds. This is in agreement since short-chain and medium-chain alkanes are generally more easily degraded due to their lower hydrophobicity. This confirms our result that degradation by consortia is more effective than individual bacteria.

CONCLUSION

Two isolates (*Acinetobacter faecalis* WD2 and *Staphylococcus* sp DD3) showed maximum growth in 1% peptone and two isolates (*Neisseria elongate* TDA4.2 and *Pseudomonas putida* TAM4.4) showed maximum growth in 0.5% peptone. Mixed bacterial consortium (ABC) gave a maximum of 98% degradation after 15 days incubation. With 100% of hydrocarbon removals were seen with the short-chain and medium chain alkanes compared to the longer chain alkanes.

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